



Investigations of antiproliferative and antioxidant activity of β -lactam morpholino-1,3,5-triazine hybrids



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ABSTRACT

This article reports for the first time the synthesis of some novel β -lactam morpholino-1,3,5-triazine hybrids by a [2+2]-cycloaddition reaction of imines **7a–c**, **9a–c** and **11** with ketenes derived from substituted acetic acids. The reaction was totally diastereoselective, leading exclusively to the formation of *cis*- β -lactams **8a–l**, **10a–f** and **12a–c**. The synthesized compounds were tested for activity towards SW1116, MCF-7 and HepG2 cancer cell lines and non-cancerous HEK-293 cell line by MTT assay. None of the compounds exert an observable effect on HepG2, MCF-7 and HEK-293 cells, but compounds **7b**, **8f**, **8g**, **8l**, **10c**, and **10e** exhibited excellent growth inhibitory activity ($IC_{50} < 5 \mu M$) against SW 1116 cells, comparable to that of doxorubicin ($IC_{50} = 6.9 \mu M$). An evaluation of the antioxidant potential of each of the compounds, performed by diphenylpicrylhydrazyl (DPPH) assay, indicated that **7b**, **9a**, **9b** and **9c** have strong free radical scavenging activity. UV absorption titration studies reveal that **7b**, **8l**, **8g** and **8f** interact strongly with calf-thymus DNA (CT-DNA) in the order of **8l** > **7b** > **8f** > **8g**. Collectively, the *in vitro* capabilities of some of these morpholino-triazine imines and β -lactams suggest possible applications to development of new antioxidants and DNA binding therapeutics.

1. Introduction

Molecular hybridization is a promising drug discovery method in which two (or more) bioactive molecules having orthogonal, complementary pharmacophoric properties are covalently linked.^{1,2} These dual- or sometimes multi-drug hybrids often improve therapeutic activity and synergism, and a decrease in the incidence of drug-resistance

versus the individual molecules.^{3–9} In this current study, we further examine an additional aspect in which we combine two important families of biologically-active compounds, namely, β -lactams (azetidine-2-ones) and 1,3,5-triazines (s-triazines), into single molecular hybrids for biological exploration. β -Lactams are an important class of compounds because of their broad biological properties (Fig. 1).^{10,11} While the most prominent β -lactams are the antibacterial agents that include

Abbreviations: MTT, Methyl Thiazol Tetrazolium bromide; IC50, Half Maximal Inhibitory Concentration; ORTEP, Oak Ridge Thermal Ellipsoid Plot; CK2, Casein kinase 2; PDB, Protein Data Bank; RMSD, Root-Mean-Square Deviation; CLogP, Calculation LogP; OD, Optical Density

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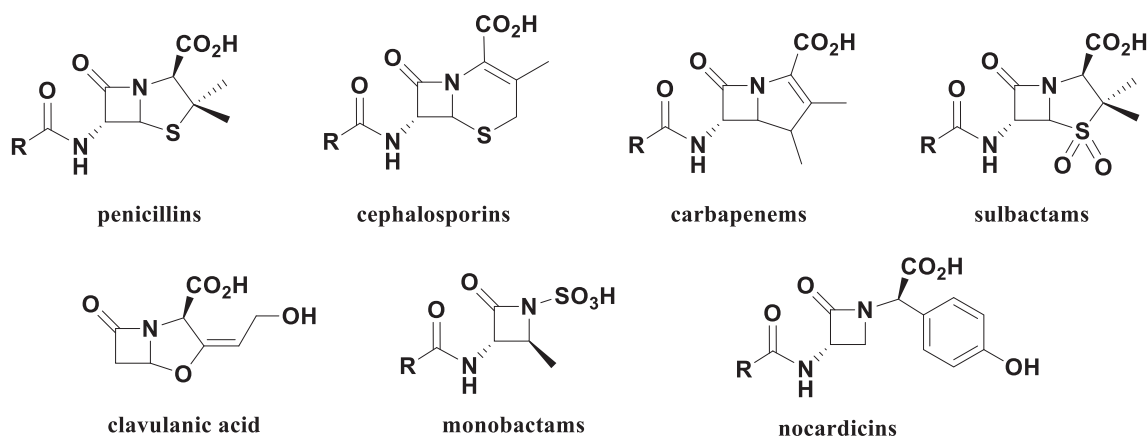
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Fig. 1. Structures of common β -lactam compounds.

the penicillins, cephalosporins, carbapenems, nocardicins, monobactams, clavulanic acid, sulbactams and tazobactams, many other β -lactams have antifungal,¹² antiviral/anti-HIV,^{13,14} anti-inflammatory,^{9,15} antimalarial,^{16,17}

or anticancer activities,^{15,18–26} and inhibit activity of human leukocyte elastases and human cytomegalovirus proteases.^{27–29} Steadily increasing levels of antibiotic resistance remains one of the important challenges in the continuing clinical utility of β -lactams.^{30,31}

Substituted *s*-triazines^{32–34} also have antimicrobial,³⁵ antiviral,³⁶ anti-inflammatory,³⁷ or anticancer activities.³⁸ Examples include the anticancer agents tretamine, furazil and dioxadet.³⁹ Bis(arylamino)-*s*-triazines are reported to be small-molecule inhibitors of anaplastic lymphoma kinase (ALK),⁴⁰ and may offer an effective treatment for anaplastic large cell lymphoma (a rare form of non-Hodgkin lymphoma), non-small cell lung cancers, and neuroblastomas in patients whose tumors contain genetic variations of ALK.⁴¹ Additionally, morpholino triazine derivatives have been reported to be inhibitors of histone deacetylases and dihydrofolate reductases,^{42,43} acting as antileukemic,⁴⁴ anticancer^{45,46} and antiproliferative agents (Fig. 2). It is widely accepted that DNA is an important target for cancer chemotherapy, and as such, the interaction of triazine compounds with DNA induces electronic perturbations that can be detected by UV–vis spectroscopy as a means to assess the strength and mode of interaction.^{43,47–53}

2. Results and discussion

2.1. Chemistry

In this study we have synthesized twenty-one new morpholino triazine- β -lactam hybrids for preliminary biological evaluation (Table 1).⁵⁴ These are divided into three series. The first group consists of β -lactams **8a–l** in which the dimorpholino triazine moiety is attached to the N-1 of the β -lactams ring. The second group consists of β -lactams **10a–f** in which the dimorpholino triazine moiety is attached to the C-4 of the β -lactams ring. The third group consists of β -lactams **12a–c** in which the dimorpholino triazine moiety is attached to both the N-1 and the C-4 centers of the β -lactam ring. The synthesis of these three groups of target compounds is outlined in (Scheme 1). The starting compound, 4,4'-(6-chloro-1,3,5-triazine-2,4-diyl) dimorpholine (**3**), was synthesized from cyanuric chloride (**1**) by a previously reported procedure.⁵⁵ A solution of cyanuric chloride in acetone and crushed ice was treated with a mixture of triethylamine and morpholine at -10°C in molar ratios of 1:4:2 to afford adduct **3**. Subsequent reaction of **3** with 4-hydroxybenzaldehyde or 4-nitroaniline in the presence of potassium carbonate and *N,N*-dimethylformamide (DMF) provided 4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy)benzaldehyde (**4**) and 4,6-dimorpholino-*N*-(4-nitrophenyl)-1,3,5-triazin-2-amine (**5**), respectively. The aryl nitro substituent of triazine **5** was reduced with Raney Ni/ N_2H_4 in

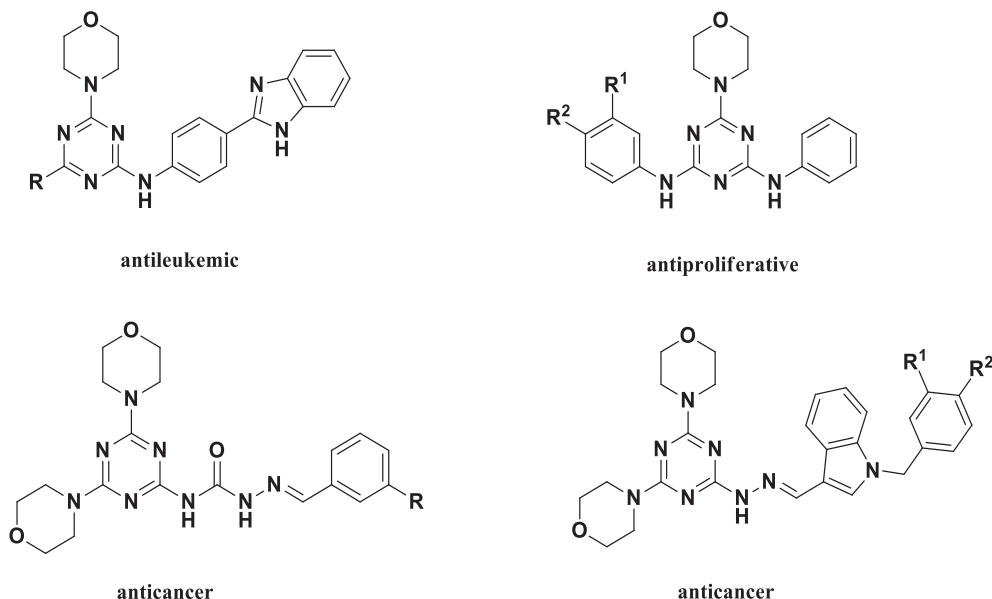
Fig. 2. Structures of morpholino-*s*-triazine derivatives with diverse biological activities.

Table 1
Structures of *cis*- β -lactam morpholino-1,3,5-triazines **8a–f**, **10a–l**, and **12a–c**.

β -lactam	Structure	Yield ^a (%)	β -lactam	Structure	Yield ^a (%)
8a		80	8f		85
8b		90	8g		80
8c		85	8h		80
8d		85	8i		80
8e		85	8j		75
8k		75	8l		75
10a		90	10d		87

(continued on next page)

Table 1 (continued)

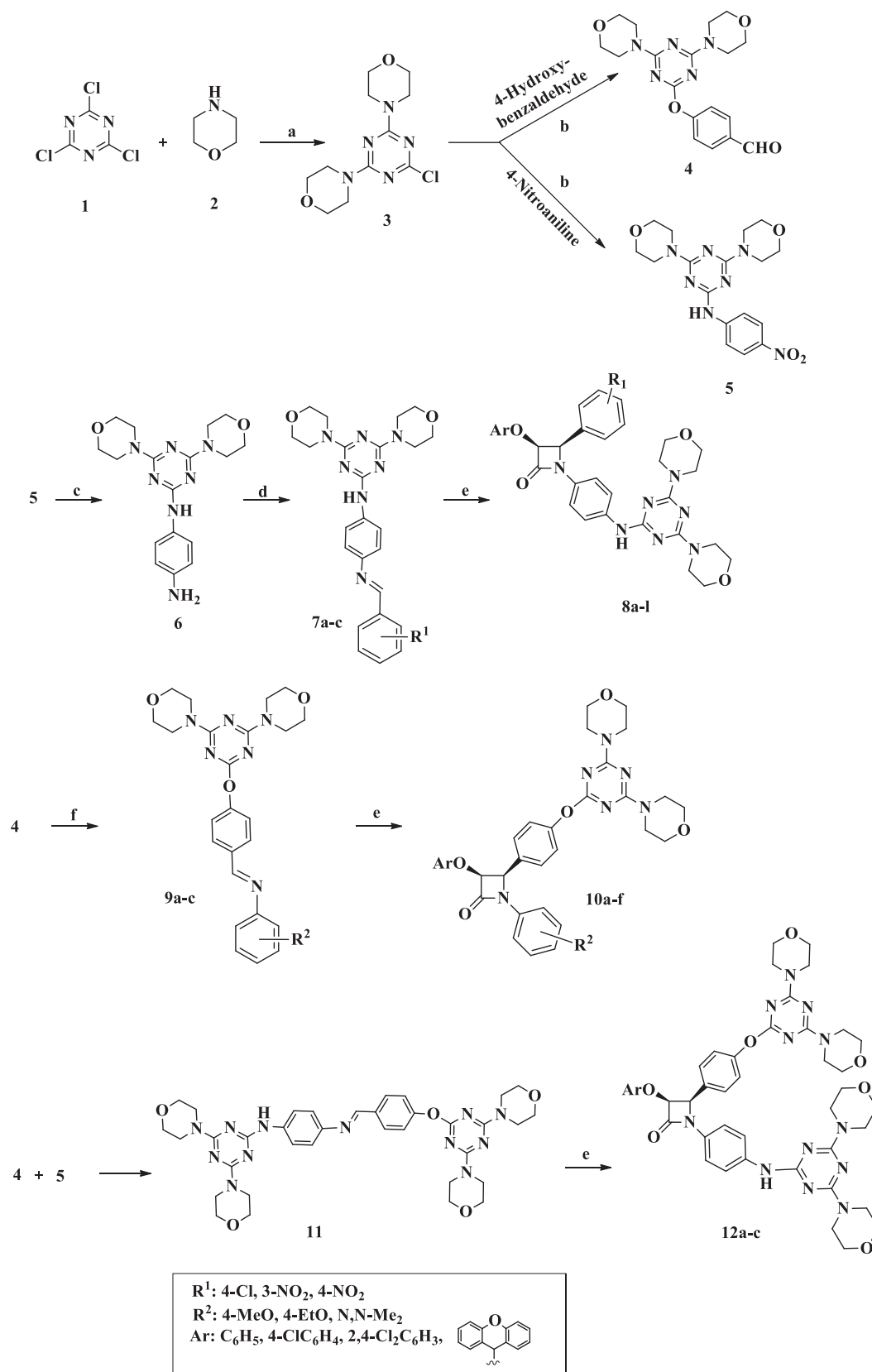
β -lactam	Structure	Yield ^a (%)	β -lactam	Structure	Yield ^a (%)
10b		85	10e		85
10c		90	10f		75
12a		70	12b		45
12c					40

^a Isolated yield.

aqueous EtOH to give 4-aminoaniline derivative **6**,⁵⁶ which reacted with various aldehydes to generate *N*-arylimines **7a–c**. Conversely, benzaldehyde **4** was reacted with various aniline derivatives to afford *N*-arylimines **9a–c**. Schiff bases **7a–c** and **9a–c** were then each combined with substituted acetic acids in the presence of triethylamine and tosyl chloride, in molar ratios of 1:1.5:5:1.5, to afford the *cis*- β -lactams **8a–l** and **10a–f** in yields of 75–90% (Scheme 1). Amine **6** and aldehyde **4** were then reacted together to prepare the Schiff base **11**, which was used to prepare *cis*- β -lactams **12a–c**.

The structures of the products were confirmed by elemental analysis and from their spectral data (IR, ¹H NMR, ¹³C NMR and mass spectra). The IR spectra of imines **7a–c**, **9a–c** and **11** showed absorption peaks at 1573–1612 cm⁻¹ due to CH=N stretching vibrations. The stretching

vibration at 1735–1759 cm⁻¹ confirmed the formation of β -lactams **8a–l**, **10a–f** and **12a–c**. The *cis* stereochemistry of the β -lactam ring substituents was assigned from the proton NMR coupling constants of the β -lactam ring hydrogens H-3 and H-4 ($J_{3,4} > 4.0$ Hz for the *cis* versus < 3.0 Hz for the *trans* stereoisomer).^{57–68} The ¹H NMR spectra of compounds **7a–c** and **9a–c** showed a singlet at δ 9.25 and 8.50 for the –NH and –N=CH, respectively. The disappearance of the –N=CH singlet for the imine **11**, and the presence of new doublets at δ 5.20–6.06 for H₃ and H₄ on the β -lactam ring, confirmed the structures of products **10a–l**, **8a–f** and **12a–c**. The ¹³C NMR spectra of each of these adducts exhibited signals at δ 43.2 and 66.7 for the two distinct CH₂ carbons of the morpholine ring, δ 55.5–62.8 for the C₃ carbon and δ 80.4–81.6 for the C₄ carbon in β -lactams **10a–l**, **8a–f** and **12a–c**. The



Scheme 1. Synthesis of novel *cis*-β-lactam morpholino-1,3,5-triazine hybrids **8a-l**, **10a-f** and **12a-c**. Reagents and conditions: (a) Et₃N, acetone; (b) K₂CO₃, reflux, DMF; (c) Raney nickel, N₂H₄, EtOH, H₂O (d) R¹PhCHO, EtOH, reflux; (e) ArCH₂CO₂H, Et₃N, CH₂Cl₂, TsCl; (f) R²PhNH₂, EtOH, reflux.

aromatic carbons also gave signals at the appropriate chemical shifts, and the β-lactam carbonyl carbon resonated at δ 161.0–162.5. Elemental and mass spectral data further confirmed the assigned product

structures. Single crystal X-ray analysis on **10f** corroborated the *cis* stereochemistry and the high planarity of the β-lactam ring (Fig. 3). The mean plane of the xanthene ring system forms dihedral angles of 90°,

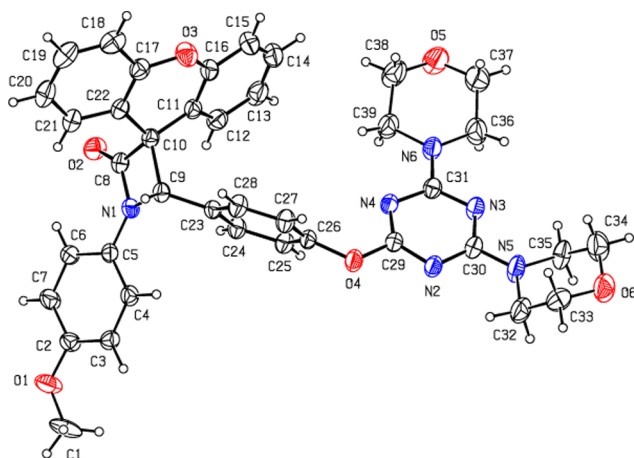


Fig. 3. ORTEP diagram of 10f.

Table 2

Hydrogen bond contact distances and geometries (Å, °) in the ORTEP structure of 10f.

D–H...A	D–H	H...A	D...A	D–H...A
C6–H6...O2	0.93	2.40	3.019 (2)	124°
C7–H7...O2	0.93	2.48	3.207 (2)	135°
C27–H27...O3	0.93	2.63	3.548 (3)	167°

102° and 125°, respectively, with the β -lactam ring and the two phenyl rings. The ORTEP image of 10f (Figure 3) indicates the presence of spatially-close C–H...O contacts (the C6–H6 to O2 contact is 2.4 Å, the C7–H7 to O2 contact is 2.48 Å, and the C27–H27 to O3 contact is 2.63 Å, listed in Table 2).^{60–64}

2.2. Antioxidant activity assay

The electron richness of *s*-triazines makes them suitable for use as potential antioxidants. Consequently, the antioxidant properties of the synthesized β -lactam-*s*-triazine hybrids were evaluated using DPPH radical scavenging assay.^{65–67} The inhibitory IC₅₀ values are displayed in Table 3. Lactams 8b and 8d were the best among the various hybrids, with IC₅₀ values of about 5 mg/ml. Lactams 10b and 10d were slightly less active, with IC₅₀ values of about 6 mg/ml. We next examined the precursor compounds, imines 7 and 9, and found surprisingly, that 9c and 9b were far more potent, with IC₅₀ values of 30 and 50 μ g/ml, respectively, about four times more effective than the positive control standard, vitamin C (IC₅₀ of 195 μ g/ml). Compounds 9a and 7b showed

Table 3

Antioxidant activity measured as % scavenging of DPPH radical.

Compounds	IC ₅₀ (μ g/ml)	Compounds	IC ₅₀ (μ g/ml)
8a	> 10000	10c	9600
8b	4800	10d	5900
8c	> 10000	10e	> 10000
8d	5800	10f	> 10000
8e	> 10000	12a	> 10000
8f	> 10000	12b	> 10000
8g	> 10000	12c	> 10000
8h	> 10000	7a	275
8i	> 10000	7b	195
8j	> 10000	7c	445
8k	> 10000	9a	195
8l	> 10000	9b	50
10a	> 10000	9c	30
10b	6200	11	> 10000
Vitamin C	195	Blank	–

Table 4

Cell growth inhibitory activity against different cancer and normal cell lines.

Compound	SW1116	IC ₅₀ (μ M)		
		HepG2	MCF-7	HEK-293
8a	17.52	>100	>100	>100
8b	>100	>100	>100	>100
8c	99.97	>100	>100	>100
8d	35.71	>100	>100	>100
8e	86.23	>100	>100	>100
8f	1.98	>100	>100	>100
8g	4.88	>100	>100	>100
8h	>100	>100	>100	>100
8i	>100	>100	>100	>100
8j	>100	>100	>100	>100
8k	>100	>100	>100	>100
8l	2.06	>100	>100	>100
10a	>100	>100	>100	>100
10b	30	>100	>100	>100
10c	2.70	>100	>100	>100
10d	>100	>100	>100	>100
10e	1.53	>100	>100	>100
10f	38.43	>100	>100	>100
12a	>100	>100	>100	>100
12b	>100	>100	>100	>100
12c	>100	>100	>100	>100
7a	93.44	>100	>100	>100
7b	3.32	>100	>100	>100
7c	15.1	>100	>100	>100
9a	>100	>100	>100	>100
9b	>100	>100	>100	>100
9c	15.64	>100	>100	>100
11	>100	>100	>100	>100
Doxorubicin (positive control)	6.9	7.3	5.5	6.1

The IC₅₀ value is defined as the concentration of a compound at which 50% cell growth inhibition is observed.

Cancer cell line: SW 1116 (colon), HepG2 (liver), MCF-7 (breast).

Normal cell line: HEK-293 (embryonic kidney).

the same IC₅₀ value as vitamin C, while imines 7a and 7c have moderate IC₅₀ values of 275 and 445 μ g/ml, respectively. The structure–activity profile for the imines 7 and 9 suggests that the compounds with an electron-releasing group offer better antioxidant properties.

2.3. Antiproliferative activity assays

As noted earlier, some *s*-triazine derivatives have anticancer properties, and thus, we next set out to examine the behavior of the β -lactam-triazine hybrids versus the normal HEK-293 (human embryonic kidney cell line) by MTT assay as well as three different cancer cell lines: SW 1116 (colon), HepG2 (liver), and MCF-7 (breast).⁶⁸ Our results are tabulated in Table 4. It is noteworthy that no inhibitory growth activity was observed for any of the 21 tested compounds towards the normal HEK-293 cells, or against the HepG2 (liver) and MCF-7 (breast) cancer cell lines at the tested concentrations (5, 10, 50, 100 and 200 μ M). This is in stark contrast to the potent activity of doxorubicin against the normal healthy (HEK-293) and cancerous (HepG2 and MCF-7) cells (IC₅₀ of 6.1, 7.3 and 5.5 μ M, respectively). On the other hand, compounds 7b, 8f, 8g, 8l, 10c and 10e demonstrated excellent inhibitory activity towards the colon cancer cells, with IC₅₀ values below 5 μ M, in comparison to the clinically-used anticancer agent, doxorubicin (IC₅₀ of 6.9 μ M). The underlying basis for the selective antiproliferative activity the synthetic compounds have towards colon cancer cells over liver or breast is unclear, but might be causally related to production of intracellular free radicals in the colon cells that induce apoptosis.

Structure-activity profiles of the β -lactams reveals that the placement of dimorpholino triazine moiety on N-1 and C-4 of the β -lactam ring enhances the antiproliferative properties. A common feature in the most active compounds is the presence of at least one chlorine atom on the phenyl rings at C-3 and C-4 of the β -lactam ring. For compounds 7b,

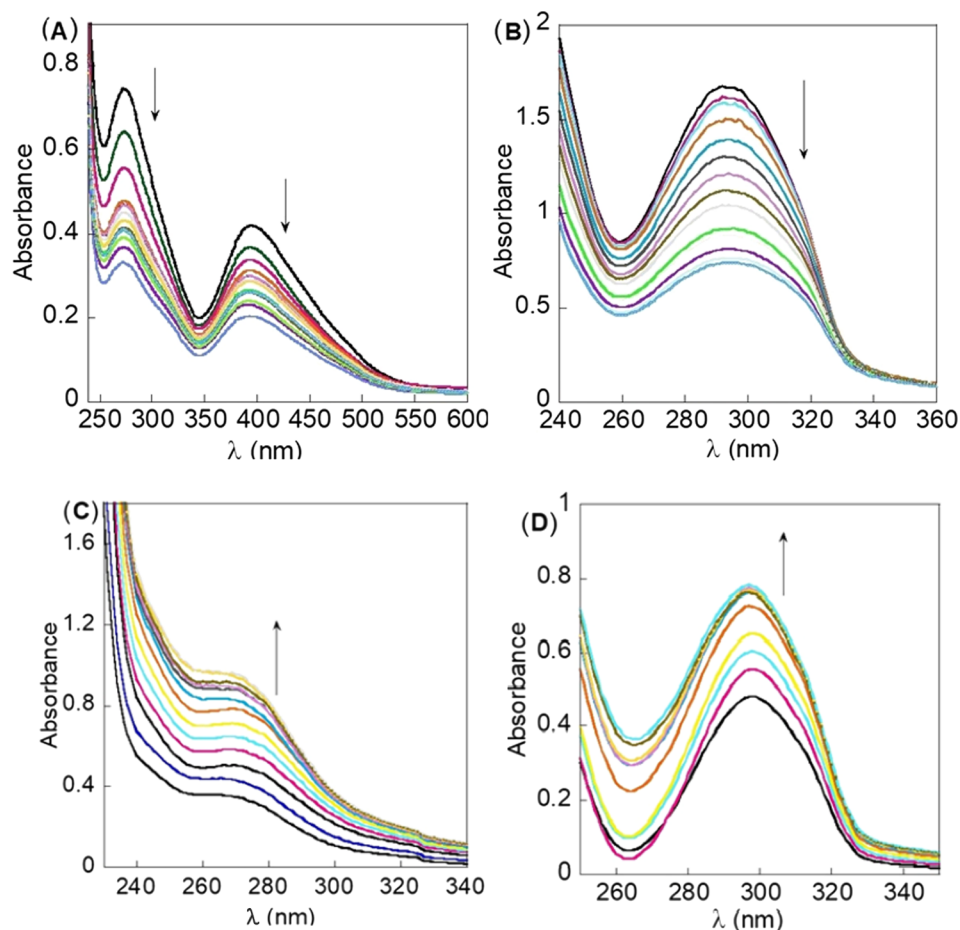


Fig. 4. Absorption titration spectra of **7b** (A), **8g** (B), **8l** (C), and **8f** (D) in 50 mM Tris HCl buffer at pH 7.4, in the absence versus presence of increasing amounts of DNA (0–22 μM **7b–8g** and 0–180 μM **8l–8f**).

8f, **8g**, **8l**, **10c** and **10e** the substituents on the aryl substituent on N-1, C-2 and C-3 of the β -lactam ring further modulate bioactivity.

2.4. UV/Vis titration studies

UV–vis absorption spectroscopy is one of the most effective techniques available to evaluate the interaction of compounds with duplex DNA. Changes in the absorbance and/or shift in wavelength position upon addition of DNA to a solution containing a fixed concentration of a test compound can give valuable information about the presence and even the type of intermolecular interaction.⁶⁹ Namely, compounds that intercalate into duplex DNA usually cause hypochromism with or without a blue or red shift. Conversely, hyperchromism in UV–vis absorption bands indicate minor groove binding, or unwinding of the DNA double helix.^{70,71} UV–vis absorption studies have been used to investigate the binding mode and strength of interaction of triazine compounds with DNA.^{43,47–53} Accordingly, UV–vis absorption titration studies were used to study the binding of complexes **7b**, **8l**, **8g** and **8f** with various concentrations of calf-thymus DNA (Fig. 4).

In the absence of CT-DNA, compounds **7b** shows absorbances at 393 and 273 nm, and **8g** has an absorbance band at 293 nm. Upon subsequent addition of CT-DNA, hypochromism was observed for both compounds, with a slight red shift of ~ 4 nm ($\lambda = 293$ nm) for complex **7b** and a slight blue shift of ~ 5 nm ($\lambda = 393$ nm) for **8g**. These observations indicate the formation of an adduct between **7b** or **8g** with CT-DNA via an intercalative mode rather than electrostatic or groove binding.⁵³ Notably, **8l** and **8f** show quite different behavior in CT-DNA titration studies. Addition of **8l** to a solution of CT-DNA produced a

hyperchromic shift of the absorption band ($\lambda = 297$ nm) along with a slight blue shift of ~ 2 nm. Complex **8f** exhibits similar behavior under similar conditions with a blue shift of ~ 2 nm ($\lambda = 268$ nm). The observed hypochromism in **8l** and **8f** with a blue shift suggested that these bind to CT-DNA by external contact, possibly electrostatic binding.⁷¹ Overall, the results suggested that although these compounds interact with CT-DNA, the mode of interaction is different for **7b** and **8g** relative to **8l** and **8f**. To compare the binding affinities of complexes with CT-DNA, quantitatively, the intrinsic binding constant K_b was calculated by monitoring the changes in their absorbance of compounds with increasing concentration of CT-DNA (see Fig. 5 and Table 5) using Eq. (1),

$$\frac{\text{DNA}}{(\epsilon_a - \epsilon_f)} = \frac{[\text{DNA}]}{(\epsilon_b - \epsilon_f)} + 1/K_b(\epsilon_b - \epsilon_f) \quad (1)$$

where [DNA] is the concentration of DNA in base pairs, ϵ_a stands for apparent extinction coefficient obtained by calculating $A_{\text{obs}}/[\text{compound}]$, ϵ_f is the extinction coefficient of each compound in their free form, and ϵ_b corresponds to the extinction coefficient of compounds in the fully bound form. The values of K_b were obtained from the ratio of the slope to intercept in the plot of $[\text{DNA}]/(\epsilon_a - \epsilon_f)$ versus [DNA]. The intrinsic binding constant values suggest a relatively strong binding to CT-DNA, and the K_b values are in the range for those of other previously reported triazine compounds. For example, Singla et al, reported the synthesis of a new series of triazine-benzimidazole analogues with different substitutions, having the intrinsic DNA binding constant K_b in the range of our compounds.^{43,44,47,48}

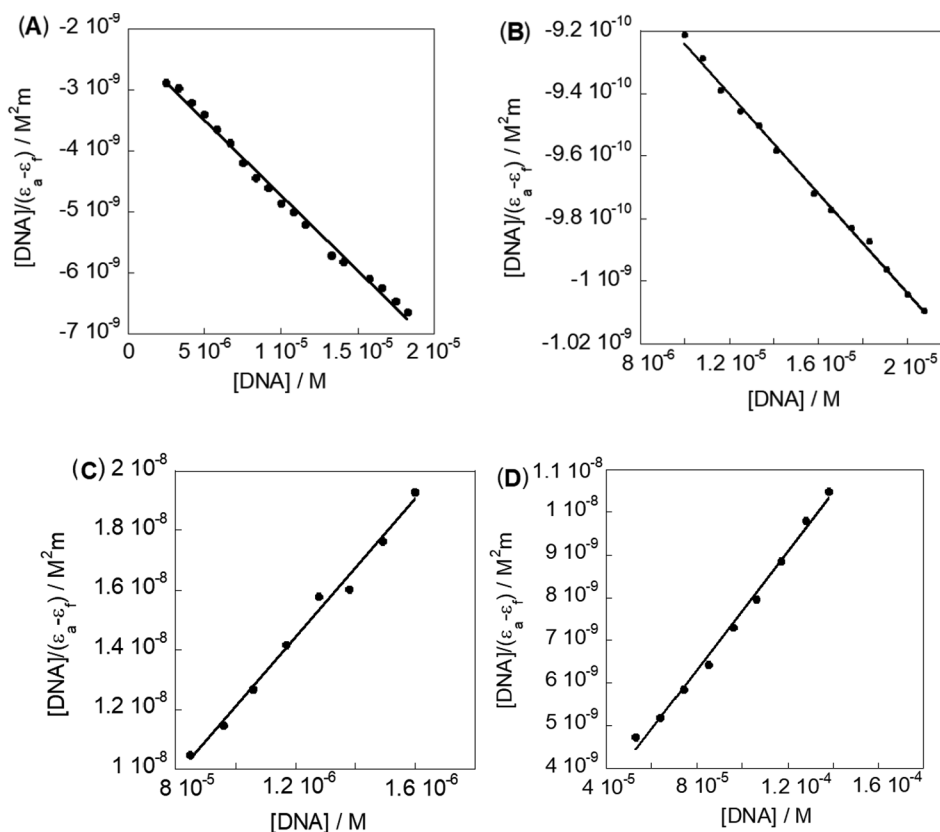


Fig. 5. Plots of $[DNA]/(\epsilon_a - \epsilon_f) / M^2 m$ vs. $[DNA] / M$ for **7b** (A), **8g** (B), **8l** (C) and **8f** (D).

Table 5

Binding constants for the interaction of **7b**, **8g**, **8l** and **8f** with CT-DNA at 25 °C in 50 mM Tris HCl buffer at pH 7.4.

Compound	λ (nm)	K_b (M^{-1})	R^{2a}
7b	393	$1.0781 \times 10^{+5}$	0.9936
8g	294	$0.9427 \times 10^{+4}$	0.9964
8l	296	$2.6000 \times 10^{+5}$	0.9942
8f	270	$0.9351 \times 10^{+5}$	0.9938

^a R^2 is the linear correlated coefficient.

3. Conclusion

In summary, a series of novel triazine-containing β -lactams hybrids were synthesized via the [2+2] ketene imine cycloaddition reaction of morpholino-1,3,5-triazine imine derivatives and different phenoxycetic acids in the presence of trimethylamine and *p*-toluenesulfonyl chloride. These cycloadditions were completely stereoselective for producing only the *cis* stereoisomers, as confirmed by proton NMR spectroscopy and X-ray crystallography. The products **7a–c**, **9a–c**, **11**, **8a–l**, **10a–f** and **12a–c** were evaluated for antioxidant and anti-proliferative activity. The compounds **9b** and **9c** showed the most potent radical scavenging activity. In the cell assays, compounds **7b**, **8f**, **8g**, **8l**, **10c**, and **10e** exhibited selective inhibitory activity towards the SW1116 colon cancer cell, with *in vitro* IC_{50} values below 5 μ M, which is comparable to that of the clinically used anticancer agent doxorubicin (IC_{50} of 6.9 μ M). Structure-activity considerations of these β -lactam-triazine hybrids suggest that the chlorophenyl group enhances *in vitro* inhibitory activity towards the colon cancer cells. UV-vis titration studies also indicated that the **7b** and **8g** are potential DNA intercalating agents, while **8f** and **8l** may bind to duplex DNA through electrostatic interactions. Additional studies are underway to further

improve the biological activity as well as to determine a better-defined mechanism of action.

4. Experimental section

4.1. General

All chemicals were purchased from Merck, Fluka and Acros chemical companies and used without further purification. Reagents and solvents such as CH_2Cl_2 and Et_3N were dried before use by distillation over CaH_2 under a nitrogen atmosphere. All known reaction products were identified by comparison of their spectral data. Infrared spectra were run on a Shimadzu FT-IR 8300 spectrophotometer using potassium bromide pellets (ν in cm^{-1}). 1H NMR and ^{13}C NMR spectra were recorded in $CDCl_3$ or dimethylsulfoxide- d_6 ($DMSO-d_6$) using a Bruker Avance DPX instrument (1H NMR at 250 MHz or 400 MHz; ^{13}C NMR at 100 MHz). Chemical shifts are reported in parts per million (δ) downfield from tetramethylsilane. Coupling constants (J) are reported in hertz (Hz). Proton signal splitting patterns are indicated as s: singlet, d: doublet, t: triplet, q: quartet, m: multiplet, dd: doublet of doublet. The mass spectra were recorded on a Shimadzu GC-MS QP 1000 EX instrument. Elemental analyses were run on a Thermo Finnigan Flash EA = 1112 series instrument. Melting points were determined on a Buchi 510 melting point apparatus and are uncorrected. Thin-layer chromatography (TLC) was carried out on silica gel 254. The crystallographic data were solved and refined by using SHELXS97 (Sheldrick, 2008) and SHELXL2018 (Sheldrick, 2015), respectively.

4.2. Preparation of 4,4'-(6-chloro-1,3,5-triazine-2,4-diyl) dimorpholine (**3**)

To a stirred solution of cyanuric chloride (**1**) (5.0 g, 27.1 mmol) in

acetone (30 ml) and crushed ice, a mixture of triethylamine (8.15 ml, 111 mmol) and morpholine (2) (4.72 ml, 54.2 mmol) was added at -10°C . After the addition, the reaction mixture was stirred at room temperature for 1 h and diluted with 50 ml of water. The white solid generated was filtered and washed with water and acetone. The white solid was dried under reduced pressure (5.79 g, 74.9% yield).

4.3. Preparation of 4,6-Dimorpholino-*N*-(4-nitrophenyl)-1,3,5-triazin-2-amine (5)

A suspension of 4,4'-(6-chloro-1,3,5-triazine-2,4-diyl) dimorpholine (300 mg, 1.05 mmol), 4-nitroaniline (159.5 mg, 1.15 mmol), and potassium carbonate (159.5 mg, 1.15 mmol), *N,N*-dimethylformamide (2 ml) was refluxed for 5 h. The precipitates generated were filtered through a Buchner funnel and washed sequentially with methanol and water. After being dried under reduced pressure, the title product (258.6 mg, 64%) was obtained as a pale yellow solid.⁵⁵

4.4. Preparation of *N*-1-(4,6-dimorpholino-1,3,5-triazin-2-yl) benzene-1,4-diamine (6)

4,6-Dimorpholino-*N*-(4-nitrophenyl)-1,3,5-triazin-2-amine (5) (710 mg, 2.2 mmol) was dissolved in 200 ml of EtOH: H₂O (9:1) by heating to reflux. The temperature was reduced to 60°C and hydrazine hydrate (250 μl , 5.15 mmol) and ca. 0.5 ml of Raney-Ni suspension were added.⁵⁶ Reflux was then maintained for 15 min. The cold mixture was filtered through a compressed pad of celite and the solvent was then evaporated. The crude product was purified by silica gel chromatography (eluent was 4–7% MeOH in CH₂Cl₂) to yield pure 6 in 83% yield.

4.4.1. *N*1-(4,6-dimorpholino-1,3,5-triazin-2-yl) benzene-1,4-diamine (6)

White solid; Mp. 265–267 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 3371 (NH₂), 1573, 1504; ¹H NMR (250 MHz, CDCl₃) δ 3.73–3.75(16H, m, 8 \times CH₂), 6.50 (1H, s, N–H), 6.64–6.68(2H, m, NH₂), 7.26–7.32(4H, m, ArH); ¹³C NMR (100 MHz, CDCl₃) δ 43.9, 67.0, 115.3, 121.9, 130.3, 141.5, 164.5, 165.3; GC–MS m/z 357.19 [M⁺]; Analysis calculated for C₁₇H₂₃N₇O₂: C, 57.13; H, 6.49; N, 27.43%. Found: C, 57.46; H, 6.12; N, 27.21%.

4.5. General procedure for the preparation of 4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy)benzaldehyde (4)

A suspension of 4,4'-(6-chloro-1,3,5-triazine-2,4-diyl) dimorpholine (300 mg, 1.05 mmol), 4-hydroxybenzaldehyde (159.5 mg, 1.15 mmol), and potassium carbonate (159.5 mg, 1.15 mmol) in *N,N*-dimethylformamide (2 ml) was refluxed for 8 h. The precipitates generated were filtered and washed sequentially with methanol and water. After being dried under reduced pressure, the title product (75%) was obtained as a pale white solid.

4.5.1. 4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy)benzaldehyde (4)

White solid; Mp. 253–255 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 2869, 1705 (CHO), 1581, 1496; ¹H NMR (250 MHz, DMSO-*d*₆) δ 3.42–3.58(16H, m, 8 \times CH₂), 7.42 (2H, d, $J = 7.5$ Hz, ArH), 7.96 (2H, d, $J = 7.5$ Hz, ArH), 9.99(1H, CHO), ¹³C NMR (100 MHz, DMSO-*d*₆) δ 43.3, 65.7, 122.4, 130.8, 133.0, 156.8, 165.4, 169.9, 191.9; GC–MS m/z 371.16 [M⁺]; Analysis calculated for C₁₈H₂₁N₅O₄: C, 58.21; H, 5.70; N, 18.86%. Found: C, 57.93; H, 5.32; N, 18.23%.

4.6. General procedure for preparation of Schiff bases 7a–c

A stirred mixture of amine 6 (1.00 mmol) and one of the aromatic aldehydes (1.00 mmol) was refluxed in ethanol containing 2–3 small drops of glacial AcOH for an appropriate time. The mixture was then cooled to room temperature, and the precipitate was filtered and recrystallized from ethanol to give Schiff bases 7a–c.

4.6.1. (*E*)-*N*-(4-((4-chlorobenzylidene) amino) phenyl)-4,6-dimorpholino-1,3,5-triazin-2-amine (7a)

Cream solid; Mp. 273–275 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 3417, 2854, 1612 (CH=N), 1542, 1504; ¹H NMR (250 MHz, DMSO-*d*₆) δ 3.60–3.69(16H, m, 8 \times CH₂), 7.26 (2H, d, $J = 10.0$ Hz, ArH), 7.55 (2H, d, $J = 7.5$ Hz, ArH), 7.71 (2H, d, $J = 7.5$ Hz, ArH), 7.90 (2H, d, $J = 7.5$ Hz, ArH), 8.66 (1H, s, CH=N), 9.24(1H, s, NH), ¹³C NMR (100 MHz, DMSO) δ 43.4, 66.1, 119.9, 121.5, 128.8, 129.8, 135.1, 135.5, 139.1, 144.2, 156.8, 163.9, 164.6; GC–MS m/z 479.18 [M⁺]; Analysis calculated for C₂₄H₂₆ClN₇O₂: C, 60.06; H, 5.46; N, 20.43%. Found: C, 60.36; H, 5.13; N, 20.23%.

4.6.2. (*E*)-4,6-dimorpholino-*N*-(4-((4-nitrobenzylidene) amino) phenyl)-1,3,5-triazin-2-amine (7b)

Orange solid; Mp. 283–285 $^{\circ}\text{C}$; IR (KBr, cm^{-1}):3417, 2854, 1612 (CH=N),1504; ¹H NMR (250 MHz, DMSO-*d*₆) δ 3.61–3.69(16H, m, 8 \times CH₂), 7.35 (2H, d, $J = 10.0$ Hz, ArH), 7.75 (2H, d, $J = 10.0$ Hz, ArH), 8.13 (2H, d, $J = 10.0$ Hz, ArH), 8.33 (2H, d, $J = 10.0$ Hz, ArH), 8.83 (1H, s, CH=N), 9.30 (1H, s, NH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 43.3, 66.0, 119.7, 121.9, 123.9, 129.3, 139.6, 142.0, 143.8, 148.4, 156.0, 163.9, 164.6; GC–MS m/z 490.21 [M⁺]; Analysis calculated for C₂₄H₂₆N₈O₄: C, 58.77; H, 5.34; N, 22.84%. Found: C, 58.24; H, 5.10; N, 23.04%.

4.6.3. (*E*)-4,6-dimorpholino-*N*-(4-((3-nitrobenzylidene) amino) phenyl)-1,3,5-triazin-2-amine (7c)

Yellow solid; Mp. 280–282 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 3417, 2854, 1612 (CH=N),1504; ¹H NMR (250 MHz, DMSO-*d*₆) δ 3.61–3.69(16H, m, 8 \times CH₂), 7.32 (2H, d, $J = 7.5$ Hz, ArH), 7.72–7.81 (3H, m, ArH), 8.29–8.33 (2H, m, ArH), 8.69 (1H, s, ArH), 8.83 (1H, s, CH=N), 9.28 (1H, s, NH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 40.9, 63.7, 117.5, 119.3, 119.9, 122.6, 128.1, 131.8, 135.5, 137.0, 141.1, 145.8, 153.6, 161.3, 162.1; GC–MS m/z 490.21 [M⁺]; Analysis calculated for C₂₄H₂₆N₈O₄: C, 58.77; H, 5.34; N, 22.84%. Found: C, 58.12; H, 5.48; N, 23.15%.

4.7. General procedure for preparation of Schiff bases 9a–c

A stirred mixture of aldehyde 5 (1.00 mmol) and one of the aniline derivatives (1.00 mmol) was refluxed in ethanol containing 2–3 small drops of glacial AcOH for an appropriate time. The mixture was then cooled to room temperature, and the precipitate was filtered and recrystallized from ethanol to give Schiff bases 9a–c.

4.7.1. (*E*)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-*N*-(4-methoxyphenyl) methanimine (9a)

Cream-colored solid; Mp. 180–182 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 2962, 2854, 1581 (CH=N),1504; ¹H NMR (250 MHz, CDCl₃) δ 3.53–3.79(16H, m, 8 \times CH₂), 3.86 (2H, s, OCH₃), 6.96 (2H, d, $J = 5.0$ Hz, ArH), 7.25–7.30 (4H, m, ArH), 7.91 (2H, d, $J = 5.0$ Hz ArH), 8.50 (1H, s, CH=N); ¹³C NMR (100 MHz, CDCl₃) δ 43.7, 55.5, 66.7, 114.4, 122.1, 122.3, 129.3, 133.2, 144.7, 154.7, 157.4, 158.3, 166.0, 170.7; GC–MS m/z 476.22 [M⁺]; Analysis calculated for C₂₅H₂₈N₆O₄: C, 63.01; H, 5.92; N, 17.64%. Found: C, 63.34; H, 5.71; N, 17.31%.

4.7.2. (*E*)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-*N*-(4-ethoxyphenyl) methanimine (9b)

Cream-colored solid; Mp. 166–168 $^{\circ}\text{C}$; IR (KBr, cm^{-1}) 2962, 2854, 1573 (CH=N), 1504; ¹H NMR (250 MHz, CDCl₃) δ 1.43 (3H, t, $J = 7.0$ Hz, CH₃), 3.60–3.83(16H, m, 8 \times CH₂), 4.06 (2H, td, $J = 6.5, 6.25$ Hz, CH₂), 6.92 (2H, d, $J = 7.5$ Hz, ArH), 7.18–7.28 (3H, m, ArH), 7.28 (1H, br, ArH), 7.89 (2H, d, $J = 8.5$ Hz, ArH), 8.48 (1H, s, CH=N); ¹³C NMR (100 MHz, CDCl₃) δ 14.8, 43.7, 63.9, 66.7, 114.9, 122.1, 122.3, 129.3, 133.3, 144.6, 154.6, 157.3, 157.7, 166.0, 170.7; GC–MS m/z 490.23 [M⁺]; Analysis calculated for C₂₆H₃₀N₆O₄: C, 63.66; H, 6.16; N, 17.13%. Found: C, 63.20; H, 6.42; N, 16.93%.

4.7.3. (E)-4-((4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)benzylidene)amino)-N,N-dimethylaniline (9c)

Green solid; Mp. 172–174 °C; IR (KBr, cm^{-1}) 2954, 2854, 1573 ($\text{CH}=\text{N}$), 1496; ^1H NMR (250 MHz, CDCl_3) δ 2.99 (6H, s, CH_3), 3.63–3.70 (16H, m, $8 \times \text{CH}_2$), 6.74 (1H, d, $J = 2.5$ Hz, ArH), 6.78 (1H, d, $J = 2.5$ Hz, ArH), 7.24 (2H, d, $J = 5.0$ Hz, ArH), 7.27 (2H, d, $J = 2.5$ Hz, ArH), 7.86 (1H, d, $J = 2.5$ Hz, ArH), 7.90 (1H, d, $J = 2.5$ Hz, ArH), 8.52 (1H, s, $\text{CH}=\text{N}$); ^{13}C NMR (100 MHz, CDCl_3) δ 40.7, 43.5, 66.7, 112.7, 122.1, 128.9, 133.6, 140.6, 149.5, 154.3, 155.3, 166.0, 170.8; GC–MS m/z 489.25 [M^+]; Analysis calculated for $\text{C}_{26}\text{H}_{31}\text{N}_7\text{O}_3$: C, 63.79; H, 6.38; N, 20.03%. Found: C, 63.98; H, 6.22; N, 19.83%.

4.8. General procedure for preparation of Schiff base 11

A stirred mixture of 4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)benzaldehyde (4) (1.00 mmol) and N1-(4,6-dimorpholino-1,3,5-triazin-2-yl)benzene-1,4-diamine (6) (1.00 mmol) was refluxed in ethanol containing 2–3 small drops of glacial AcOH for an appropriate time. The mixture was then cooled to room temperature, and the precipitate was filtered and recrystallized from ethanol to give Schiff base 11.

4.8.1. (E)-N-(4-((4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)benzylidene)amino)phenyl)-4,6-dimorpholino-1,3,5-triazin-2-amine (11)

Lemon yellow solid; Mp. 252–254 °C; IR (KBr, cm^{-1}) 2923, 2854, 1542 ($\text{CH}=\text{N}$), 1496; ^1H NMR (400 MHz, CDCl_3) δ 3.63 (16H, d, $J = 4.0$ Hz, $8 \times \text{CH}_2$), 3.66 (16H, d, $J = 4.0$ Hz, $8 \times \text{CH}_2$), 6.71 (1H, s, NH), 7.16 (1H, s, ArH), 7.18 (2H, d, $J = 4.0$ Hz, ArH), 7.21 (1H, s, ArH), 7.51 (2H, d, $J = 8.0$ Hz, ArH), 7.83 (2H, d, $J = 8.0$ Hz, ArH), 8.44 (1H, s, $\text{CH}=\text{N}$); ^{13}C NMR (100 MHz, DMSO) δ 43.1, 66.0, 101.6, 120.0, 121.3, 122.1, 129.3, 133.1, 138.8, 154.2, 157.3, 163.9, 164.6, 165.4, 170.0; GC–MS m/z 710.34 [M^+]; Analysis calculated for $\text{C}_{35}\text{H}_{42}\text{N}_{12}\text{O}_5$: C, 59.14; H, 5.96; N, 23.65%. Found: C, 58.98; H, 6.12; N, 23.93%.

4.9. General procedure for the synthesis of novel di morpholino-1,3,5-triazin β -lactam hybrids 8a–l, 10a–f and 12a–c

A mixture of Schiff bases 7a–c or 9a–b or 11 (1.00 mmol), triethylamine (5.00 mmol), a substituted acetic acid (1.50 mmol) and tosyl chloride (1.50 mmol) in dry CH_2Cl_2 (15 ml) was stirred overnight at room temperature. Then the mixture was washed with 1 N aqueous HCl (20 ml), saturated aqueous NaHCO_3 (20 ml) and brine (20 ml). The organic layer was dried (anhydrous Na_2SO_4), filtered and evaporated to give crude products 8a–l or 10a–f and 12a–c that were purified by recrystallization from CH_2Cl_2 .

4.9.1. 1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(4-nitrophenyl)-3-phenoxyazetid-2-one (8a)

White solid; Mp. 228–230 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751, (CO β -lactam), 1566, 1512; ^1H NMR (250 MHz, CDCl_3) δ 3.60–3.68 (16H, m, $8 \times \text{CH}_2$), 5.47 (1H, d, $J = 5.0$ Hz, H-4), 5.63 (1H, d, $J = 5.0$ Hz, H-3), 6.71 (1H, s, NH), 6.74 (1H, s, ArH), 6.80 (1H, d, $J = 7.5$ Hz, ArH), 6.91 (1H, t, $J = 7.5$ Hz, ArH), 7.05 (1H, t, $J = 10.0$ Hz, ArH), 7.14 (2H, t, $J = 7.5$ Hz, ArH), 7.24–7.30 (2H, m, ArH), 7.43–7.53 (3H, m, ArH), 8.11 (2H, d, $J = 7.5$ Hz, ArH); ^{13}C NMR (100 MHz, DMSO) δ 43.3, 60.0, 65.7, 80.8, 114.3, 117.21, 120.1, 120.7, 122.1, 123.2, 129.4, 136.0, 140.9, 147.4, 157.7, 162.3, 164.1, 165.6; GC–MS m/z 624.24 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{32}\text{N}_8\text{O}_6$: C, 61.53; H, 5.16; N, 17.94%. Found: C, 61.01; H, 5.82; N, 17.94%.

4.9.2. 4-(4-chlorophenyl)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-3-phenoxyazetid-2-one (8b)

White solid; Mp. 271–273 °C; IR (KBr, cm^{-1}): 3402, 2854, 1759 (CO β -lactam), 1366, 1504; ^1H NMR (250 MHz, CDCl_3) δ 3.70–3.77 (16H, m, $8 \times \text{CH}_2$), 5.36 (1H, d, $J = 5.0$ Hz, H-4), 5.57 (1H, d, $J = 5.0$ Hz, H-3),

6.67 (1H, s, NH), 6.78 (2H, d, $J = 7.5$ Hz, ArH), 6.94 (1H, t, $J = 7.5$ Hz, ArH), 7.21 (2H, t, $J = 7.5$ Hz, ArH), 7.25–7.34 (6H, m, ArH), 7.47 (2H, d, $J = 7.5$ Hz, ArH); ^{13}C NMR (100 MHz, CDCl_3) δ 43.6, 61.4, 66.6, 81.1, 115.5, 117.9, 120.2, 122.3, 128.7, 129.3, 129.4, 131.3, 131.4, 134.6, 136.3, 156.7, 162.3, 164.1, 165.1; GC–MS m/z 613.22 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{32}\text{ClN}_7\text{O}_4$: C, 62.59; H, 5.25; N, 15.97%. Found: C, 62.09; H, 5.62; N, 16.13%.

4.9.3. 3-(4-chlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(3-nitrophenyl) azetid-2-one (8c)

White solid; Mp. 268–270 °C; IR (KBr, cm^{-1}) 3417, 2854, 1759 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO}-d_6$) δ 3.60–3.64 (16H, m, $8 \times \text{CH}_2$), 5.91 (1H, d, $J = 5.0$, H-4), 5.96 (1H, d, $J = 5.0$, H-3) 6.75–6.85 (2H, m, ArH), 7.23 (4H, t, $J = 10.0$ Hz, ArH), 7.57 (1H, t, $J = 7.5$ Hz, ArH), 7.63 (2H, d, $J = 9.0$ Hz, ArH), 7.77 (1H, d, $J = 7.5$ Hz, ArH), 8.09 (1H, d, $J = 7.5$ Hz, ArH), 8.19 (1H, s, ArH), 9.15 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 43.2, 59.5, 65.9, 80.4, 116.7, 114.9, 117.4, 120.1, 122.8, 123.3, 125.7, 128.6, 129.1, 130.2, 134.4, 135.6, 137.2, 147.3, 154.7, 161.4, 163.8, 164.6; GC–MS m/z 658.21 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{31}\text{ClN}_8\text{O}_6$: C, 58.31; H, 4.74; N, 17.00%. Found: C, 57.91; H, 4.13; N, 17.31%.

4.9.4. 3-(2,4-dichlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(4-nitrophenyl) azetid-2-one (8d)

White solid; Mp. 190–192 °C; IR (KBr, cm^{-1}) 3440, 2854, 1759 (CO β -lactam), 1620, 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO}-d_6$) δ 3.58–3.63 (16H, m, $8 \times \text{CH}_2$), 5.90 (1H, d, $J = 5.0$ Hz, H-4), 6.06 (1H, d, $J = 5.0$ Hz, H-3), 6.79 (1H, d, $J = 7.5$ Hz, ArH), 7.21 (3H, d, $J = 7.5$ Hz, ArH), 7.36–7.43 (1H, m, ArH), 7.50–7.75 (4H, m, ArH), 8.13 (1H, d, $J = 10.0$ Hz, ArH), 9.15 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 43.2, 59.5, 66.0, 81.0, 115.0, 116.3, 117.4, 120.1, 123.2, 126.0, 127.3, 127.9, 128.6, 129.3, 130.1, 137.3, 140.8, 147.4, 161.0, 163.8, 164.6; GC–MS $m/z = 692.17$ [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{30}\text{Cl}_2\text{N}_8\text{O}_6$: C, 55.42; H, 4.36; N, 16.16%. Found: C, 55.01; H, 4.64; N, 15.91%.

4.9.5. 4-(4-chlorophenyl)-3-(2,4-dichlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl) azetid-2-one (8e)

White solid; Mp. 275–277 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO}-d_6$) δ 3.59–3.63 (16H, m, $8 \times \text{CH}_2$), 5.71 (1H, d, $J = 4$ Hz, H-4), 5.97 (1H, d, $J = 4$ Hz, H-3), 7.20 (3H, d, $J = 7.5$ Hz, ArH), 7.30–7.44 (6H, m, ArH), 7.62 (2H, d, $J = 7.5$ Hz, ArH), 9.14 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 44.5, 60.8, 67.2, 81.9, 117.3, 118.7, 121.3, 123.4, 127.1, 120.0, 129.4, 130.6, 131.1, 131.6, 133.1, 134.2, 138.4, 151.8, 162.3, 165.0, 165.8; GC–MS m/z 681.14 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{30}\text{Cl}_3\text{N}_7\text{O}_4$: C, 56.28; H, 4.43; N, 14.36%. Found: C, 56.71; H, 4.23; N, 13.96%.

4.9.6. 1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(3-nitrophenyl)-3-phenoxyazetid-2-one (8f)

White solid; Mp. 233–235 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751 (CO β -lactam) 1612, 1566, 1527; ^1H NMR (250 MHz, $\text{DMSO}-d_6$) δ 3.34–3.75 (16H, m, $8 \times \text{CH}_2$), 5.33 (2H, br, H-3, H-4), 6.78 (2H, d, $J = 7.5$ Hz, ArH), 6.86 (1H, t, $J = 7.5$ Hz, ArH), 6.91 (1H, d, $J = 7.5$ Hz, ArH), 7.11–7.16 (2H, m, ArH), 7.20 (1H, d, $J = 4.5$ Hz, ArH), 7.25 (1H, d, $J = 7.5$ Hz, ArH), 7.33 (1H, d, $J = 7.5$ Hz, ArH), 7.55 (1H, t, $J = 7.5$ Hz, ArH), 7.79 (1H, d, $J = 7.5$ Hz, ArH), 8.06–8.10 (1H, m, ArH), 8.21 (1H, s, ArH), 9.14 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 43.2, 59.7, 65.7, 80.5, 114.3, 114.9, 117.2, 120.7, 122.0, 123.4, 129.4, 134.4, 134.9, 136.1, 147.3, 155.8, 157.7, 162.4, 164.1, 165.6; GC–MS m/z 624.24 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{32}\text{N}_8\text{O}_6$: C, 61.53; H, 5.16; N, 17.94%. Found: C, 61.09; H, 5.62; N, 17.33%.

4.9.7. 3-(4-chlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(4-nitrophenyl) azetidin-2-one (8g)

Cream solid; Mp. 218–220 °C; IR (KBr, cm^{-1}) 3417, 2854, 1759 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.59–3.63(16H, m, $8 \times \text{CH}_2$), 5.72 (1H, d, $J = 4.2$ Hz, H-4), 5.97 (1H, d, $J = 4.2$ Hz, H-3), 7.20 (4H, d, $J = 7.5$ Hz ArH), 7.30–7.49 (6H, m, ArH), 7.62 (2H, d $J = 7.5$ Hz, ArH), 9.14 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.2, 59.7, 65.7, 80.5, 114.3, 114.9, 117.2, 120.7, 122.0, 123.4, 129.4, 134.4, 134.9, 136.1, 147.3, 155.8, 157.7, 162.4, 164.1, 165.6; GC-MS m/z 658.21 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{31}\text{ClN}_8\text{O}_6$: C, 58.31; H, 4.74; N, 17.00%. Found: C, 57.96; H, 4.01; N, 17.43%.

4.9.8. 3-(4-chlorophenoxy)-4-(4-chlorophenyl)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl) azetidin-2-one (8h)

White solid; Mp. 208–210 °C; IR (KBr, cm^{-1}) 3417, 2854, 1759 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (250 MHz, CDCl_3) δ 3.64–3.73(16H, m, $8 \times \text{CH}_2$), 5.48 (1H, d, $J = 5.0$ Hz, H-4), 5.58 (1H, d, $J = 5.0$ Hz, H-3), 6.69 (1H, s, N-H), 6.74 (2H, d, $J = 10$ Hz, ArH), 7.14 (2H, d, $J = 7.5$ Hz, ArH), 7.24–7.27 (3H, m, ArH), 7.49 (2H, d, $J = 10.0$ Hz, ArH), 7.55 (2H, d, $J = 10.0$ Hz, ArH) 8.18 (1H, d, $J = 4.5$ Hz, ArH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.3, 60.2, 63.1, 65.7, 80.6, 114.9, 116.7, 118.5, 121.6, 125.5, 128.9, 129.0, 129.5, 129.7, 152.0, 155.2, 155.3, 161.5, 165.3, 170.3; GC-MS m/z 647.18 [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{31}\text{Cl}_2\text{N}_7\text{O}_4$: C, 59.26; H, 4.82; N, 15.12%. Found: C, 59.41; H, 4.32; N, 15.59%.

4.9.9. 3-(2,4-dichlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-4-(3-nitrophenyl) azetidin-2-one (8i)

White solid; Mp. 273–275 °C; IR (KBr, cm^{-1}) 3417, 2854, 1759 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.59–3.63(16H, m, $8 \times \text{CH}_2$), 5.91 (1H, d, $J = 5.0$ Hz, H-4), 6.08 (1H, d, $J = 5.0$ Hz, H-3) 7.24–7.33 (4H, m, ArH), 7.35–7.39 (1H, m, ArH), 7.54 (1H, t, $J = 8$ Hz, H-3), 7.64 (2H, d, $J = 10$ Hz, ArH), 7.76 (1H, d, $J = 7.5$ Hz, ArH), 8.09–8.12 (1H, m, ArH), 8.18 (1H, s, ArH), 9.15 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.2, 59.1, 65.9, 80.0, 116.1, 117.4, 120.1, 122.0, 122.7, 129.3, 129.8, 130.1, 134.5, 135.2, 137.3, 147.3, 150.2, 161.0, 163.8, 164.6; GC-MS $m/z = 692.17$ [M^+]; Analysis calculated for $\text{C}_{32}\text{H}_{30}\text{Cl}_2\text{N}_8\text{O}_6$: C, 55.42; H, 4.36; N, 16.16%. Found: C, 55.01; H, 4.64; N, 15.91%.

4.9.10. 1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-2-(4-nitrophenyl) spiro[azetidine-3,9'-xanthen]-4-one (8j)

White solid; Mp. 283–285 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751 (CO β -lactam), 1604, 1573, 1512; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.43–3.66(16H, m, $8 \times \text{CH}_2$), 5.55 (1H, s, H-4), 6.83 (1H, t, $J = 7.0$ Hz, ArH), 7.02 (1H, d, $J = 7.5$ Hz, ArH), 7.08–7.16 (2H, m, ArH), 7.23 (2H, d, $J = 7.5$ Hz, ArH), 7.30 (2H, d, $J = 10.0$ Hz, ArH), 7.36–7.50 (4H, m, ArH), 7.72 (2H, d, $J = 10.0$ Hz, ArH), 7.95 (2H, d, $J = 7.5$ Hz, ArH), 9.21 (1H, s, NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.2, 63.2, 65.9, 72.4, 115.9, 116.1, 116.79, 118.0, 120.1, 120.4, 122.6, 122.8, 123.1, 124.7, 125.7, 127.1, 128.1, 128.5, 129.6, 129.7, 130.3, 137.4, 141.6, 146.6, 150.9, 151.4, 163.8, 164.4, 164.6; GC-MS m/z 698.26 [M^+]; Analysis calculated for $\text{C}_{38}\text{H}_{34}\text{N}_8\text{O}_6$: C, 65.32; H, 4.90; N, 16.04%. Found: C, 65.71; H, 4.21; N, 16.52%.

4.9.11. 1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl)-2-(3-nitrophenyl) spiro[azetidine-3,9'-xanthen]-4-one (8k)

White solid; Mp. 269–271 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751 (CO β -lactam), 1573, 1514; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.59–3.65(16H, m, $8 \times \text{CH}_2$), 5.59 (1H, s, H-4), 6.87–7.09 (6H, m, ArH), 7.30–7.42 (7H, m, ArH), 7.73 (2H, s, ArH), 7.92 (1H, s, ArH), 9.21 (1H, s, NH); ^{13}C NMR (100 MHz, DMSO) δ 43.2, 63.1, 65.9, 71.9, 116.5, 117.9, 120.1, 121.2, 122.6, 122.8, 123.5, 124.7, 125.4, 125.7, 127.1, 128.0, 128.3, 129.7, 130.1, 130.3, 133.3, 136.3, 137.4, 147.1, 151.0, 151.5, 163.8, 164.3, 164.6; GC-MS m/z 698.26 [M^+]; Analysis calculated for

$\text{C}_{38}\text{H}_{34}\text{N}_8\text{O}_6$: C, 65.32; H, 4.90; N, 16.04%. Found: C, 65.61; H, 4.38; N, 15.74%.

4.9.12. 2-(4-chlorophenyl)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) amino) phenyl) spiro[azetidine-3,9'-xanthen]-4-one (8l)

White solid; Mp. 273–275 °C; IR (KBr, cm^{-1}) 3417, 2854, 1751 (CO β -lactam), 1504; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.42–3.67(16H, m, $8 \times \text{CH}_2$), 5.33 (1H, s, H-4), 6.87 (1H, d, $J = 5$ Hz, ArH), 6.95 (4H, t, $J = 7.5$ Hz, ArH), 7.02 (1H, d, $J = 7.5$ Hz, ArH), 7.08 (1H, d, $J = 7.5$ Hz, ArH), 7.12–7.17 (2H, m, ArH), 7.25 (2H, t, $J = 8.5$ Hz, ArH), 7.33–7.39 (3H, m, ArH), 7.45 (1H, d, $J = 7.5$ Hz, ArH), 7.70 (1H, d, $J = 7.5$ Hz, ArH), 9.20 (1H, s, N-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.3, 62.8, 65.9, 72.6, 116.5, 116.8, 118.0, 120.1, 120.8, 122.6, 122.7, 123.5, 124.6, 125.4, 125.7, 127.1, 128.0, 128.5, 129.4, 129.6, 130.1, 132.2, 132.7, 137.2, 151.2, 163.8, 164.5, 164.6; GC-MS $m/z = 687.24$ [M^+]; Analysis calculated for $\text{C}_{38}\text{H}_{34}\text{ClN}_7\text{O}_4$: C, 66.32; H, 4.98; N, 14.25%. Found: C, 66.07; H, 5.04; N, 14.61%.

4.9.13. 4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-1-(4-methoxyphenyl)-3-phenoxyazetidin-2-one (10a)

White solid; Mp. 193–195 °C; IR (KBr, cm^{-1}) 2962, 2916, 2854, 1759 (CO β -lactam), 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.32–3.66(16H, m, $8 \times \text{CH}_2$), 3.68 (3H, s, OCH_3), 5.71 (1H, d, $J = 0.5$ Hz, H-4), 5.81 (1H, d, $J = 0.5$ Hz, H-3) 6.81 (2H, d, $J = 7.5$ Hz, ArH), 6.88–6.93 (3H, m, ArH), 7.03 (2H, d, $J = 7.5$ Hz, ArH), 7.16 (2H, d, $J = 7.5$ Hz, ArH), 7.20 (1H, d, $J = 4.5$ Hz, ArH), 7.25 (1H, d, $J = 7.5$ Hz, ArH), 7.33 (1H, d, $J = 7.5$ Hz, ArH), 7.23 (2H, d, $J = 10.0$ Hz, ArH), 7.37 (2H, d, $J = 10.0$ Hz, ArH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.3, 55.2, 60.4, 65.7, 80.6, 114.4, 115.1, 118.4, 121.6, 128.9, 129.3, 129.7, 129.9, 152.0, 155.9, 156.5, 162.0, 165.4, 170.3; GC-MS m/z 610.25 [M^+]; Analysis calculated for $\text{C}_{33}\text{H}_{34}\text{N}_6\text{O}_6$: C, 64.91; H, 5.61; N, 13.76%. Found: C, 65.12; H, 5.83; N, 13.36%.

4.9.14. 3-(4-chlorophenoxy)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-1-(4-methoxyphenyl) azetidin-2-one (10b)

White solid; Mp. 203–205 °C; IR (KBr, cm^{-1}) 2962, 2854, 1743 (CO β -lactam), 1581; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 3.33–3.62(16H, m, $8 \times \text{CH}_2$), 3.76 (3H, s, OCH_3), 5.70 (1H, d, $J = 4.5$ Hz, H-4), 5.81 (1H, d, $J = 4.5$ Hz, H-3), 6.86 (4H, t, $J = 10.0$ Hz, ArH), 7.20–7.24 (4H, m, ArH), 7.35 (2H, d, $J = 7.5$ Hz, ArH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 43.2, 55.2, 60.2, 65.7, 80.6, 114.5, 116.7, 118.5, 122.6, 125.5, 128.9, 129.0, 129.5, 129.9, 152.0, 155.2, 155.9, 161.5, 165.3, 170.3; GC-MS m/z 644.22 [M^+]; Analysis calculated for $\text{C}_{33}\text{H}_{33}\text{ClN}_6\text{O}_6$: C, 61.44; H, 5.16; N, 13.03%. Found: C, 61.12; H, 5.42; N, 13.35%.

4.9.15. 3-(2,4-dichlorophenoxy)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-1-(4-methoxyphenyl) azetidin-2-one (10c)

White solid; Mp. 269–267 °C; IR (KBr, cm^{-1}) 2962, 2923, 2854, 1735 (CO β -lactam), 1620, 1573, 1504; ^1H NMR (250 MHz, CDCl_3) δ 3.47–3.64(16H, m, $8 \times \text{CH}_2$), 3.76 (3H, s, OCH_3), 5.40 (1H, d, $J = 5.0$ Hz, H-4), 5.49 (1H, d, $J = 5.0$ Hz, H-3) 6.81 (2H, d, $J = 10.0$ Hz, ArH), 7.12–7.23 (5H, m, ArH), 7.31 (2H, d, $J = 10.0$ Hz, ArH), 7.40 (2H, d, $J = 10.0$ Hz, ArH); ^{13}C NMR (100 MHz, CDCl_3) δ 43.7, 55.4, 61.1, 66.6, 81.8, 114.4, 116.8, 118.9, 122.1, 125.4, 127.5, 127.6, 128.8, 128.9, 129.9, 130.1, 151.6, 152.9, 156.7, 161.7, 166.0, 170.7; GC-MS m/z 678.18 [M^+]; Analysis calculated for $\text{C}_{33}\text{H}_{32}\text{Cl}_2\text{N}_6\text{O}_6$: C, 58.33; H, 4.75; N, 12.37%. Found: C, 58.69; H, 4.32; N, 12.73%.

4.9.16. 4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl) oxy) phenyl)-1-(4-ethoxyphenyl)-3-phenoxyazetidin-2-one (10d)

White solid; Mp. 218–220 °C; IR (KBr, cm^{-1}) 2970, 2908, 2854, 1759 (CO β -lactam), 1573, 1504; ^1H NMR (250 MHz, $\text{DMSO-}d_6$) δ 1.26 (3H, t, $J = 0.75$ Hz, OCH_3) 3.34–3.62(16H, m, $8 \times \text{CH}_2$), 3.93 (2H, q, $J = 7.0$ Hz, CH_2), 5.70 (1H, d, $J = 4.8$ Hz, H-4), 5.80 (1H, d, $J = 4.8$ Hz, H-3) 6.80 (2H, d, $J = 7.5$ Hz, ArH), 6.81–6.93 (3H, m, ArH), 7.03 (2H, d, $J = 10.0$ Hz, ArH), 7.15–7.23 (4H, m, ArH), 7.36

(2H, d, $J = 7.5$ Hz ArH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 14.55, 43.1, 60.4, 63.1, 65.7, 80.6, 114.9, 115.0, 118.4, 121.6, 121.8, 128.9, 129.2, 129.7, 129.8, 152.0, 155.2, 156.5, 162.0, 165.3, 170.3; GC-MS m/z 624.27 [M^+]; Analysis calculated for $\text{C}_{34}\text{H}_{36}\text{N}_6\text{O}_6$: C, 65.37; H, 5.81; N, 13.45%. Found: C, 65.11; H, 5.32; N, 13.78%.

4.9.17. 3-(4-chlorophenoxy)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)phenyl)-1-(4-ethoxyphenyl) azetidin-2-one (10e)

White solid; Mp. 204–206 °C; IR (KBr, cm^{-1}) 2923, 1743 (CO β -lactam), 1589, 1496; ^1H NMR (250 MHz, DMSO- d_6) δ 1.26 (3H, t, $J = 7.5$ Hz, OCH_3), 3.34–3.82 (16H, m, $8 \times \text{CH}_2$), 3.93 (2H, q, $J = 7.5$ Hz, CH_2), 5.71 (1H, d, $J = 5.0$ Hz, H-4), 5.82 (1H, d, $J = 5.0$ Hz, H-3), 6.83–6.90 (4H, m, ArH), 7.04 (2H, d, $J = 7.5$ Hz, ArH), 7.19–7.25 (4H, m, ArH), 7.36 (2H, d, $J = 10.0$ Hz ArH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 14.55, 43.2, 60.2, 63.1, 65.7, 80.6, 114.9, 116.7, 118.5, 121.6, 125.5, 128.9, 129.5, 129.7, 152.0, 155.2, 155.3, 161.5, 165.3, 170.3; GC-MS m/z 658.23 [M^+]; Analysis calculated for $\text{C}_{34}\text{H}_{35}\text{ClN}_6\text{O}_6$: C, 61.96; H, 5.35; N, 12.75%. Found: C, 62.1; H, 5.63; N, 12.23%.

4.9.18. 2-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)phenyl)-1-(4-methoxyphenyl) spiro[azetidine-3,9'-xanthen]-4-one (10f)

White solid; Mp. 232–234 °C; IR (KBr, cm^{-1}) 2970, 2900, 2854, 1751 (CO β -lactam), 1573, 1504; ^1H NMR (250 MHz, DMSO- d_6) δ 3.43–3.71 (16H, m, $8 \times \text{CH}_2$), 3.76 (3H, s, OCH_3), 5.35 (1H, s, H-4), 6.91 (3H, d, $J = 5.0$ Hz), 6.99 (4H, t, $J = 5.0$ Hz, ArH), 7.05 (1H, d, $J = 5.0$ Hz, ArH), 7.15 (1H, d, $J = 7.5$ Hz, ArH), 7.20 (1H, d, $J = 7.5$ Hz, ArH), 7.30 (2H, d, $J = 7.5$ Hz, ArH), 7.35 (1H, d, $J = 4.5$ Hz, ArH), 7.43 (2H, d, $J = 5.0$ Hz, ArH), 7.47 (1H, d, $J = 4.8$ Hz, ArH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 43.3, 55.3, 62.8, 65.7, 73.0, 114.5, 115.9, 116.6, 119.2, 121.1, 121.3, 122.7, 124.5, 125.7, 127.5, 128.8, 129.1, 129.5, 129.9, 151.0, 151.4, 151.5, 156.2, 164.7, 165.4, 170.0; GC-MS m/z 684.27 [M^+]; Analysis calculated for $\text{C}_{39}\text{H}_{36}\text{N}_6\text{O}_6$: C, 68.41; H, 5.30; N, 12.27%. Found: C, 68.74; H, 5.12; N, 12.67%.

4.9.19. 1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)amino)phenyl)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)phenyl)-3-phenoxyazetidin-2-one (12a)

White solid; Mp. 263–265 °C; IR (KBr, cm^{-1}) 2854, 1759 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (400 MHz, DMSO- d_6) δ 3.55–3.62 (16H, m, $8 \times \text{CH}_2$), 3.66–3.67 (16H, m, $8 \times \text{CH}_2$), 5.72 (1H, d, $J = 4.8$ Hz, H-4), 5.83 (1H, d, $J = 4.8$ Hz, H-3), 6.83 (2H, d, $J = 8.0$ Hz, ArH), 6.92 (1H, t, $J = 7.6$ Hz, ArH), 7.05 (2H, d, $J = 12.0$ Hz, ArH), 7.21 (4H, t, $J = 8.8$ Hz, ArH), 7.37 (2H, d, $J = 8.8$ Hz, ArH), 7.64 (2H, d, $J = 8.8$ Hz, ArH), 9.18 (1H, s, NH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 43.2, 60.2, 65.9, 80.4, 115.0, 117.3, 119.9, 121.6, 121.8, 126.7, 128.9, 129.3, 136.8, 151.9, 152.9, 156.5, 162.0, 163.8, 165.3, 170.3; GC-MS m/z 844.38 [M^+]; Analysis calculated for $\text{C}_{43}\text{H}_{48}\text{N}_{12}\text{O}_7$: C, 61.13; H, 5.73; N, 19.89%. Found: C, 60.84; H, 5.13; N, 19.27%.

4.9.20. 3-(4-chlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)amino)phenyl)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)phenyl) azetidin-2-one (12b)

White solid; Mp. 275–277 °C; IR (KBr, cm^{-1}) 2854, 1751 (CO β -lactam), 1573, 1504; ^1H NMR (400 MHz, CDCl_3) δ 3.73 (16H, d, $J = 4.4$ Hz, $8 \times \text{CH}_2$), 3.75 (16H, d, $J = 4.8$ Hz, $8 \times \text{CH}_2$), 5.45 (1H, d, $J = 4.8$ Hz, H-4), 5.51 (1H, d, $J = 4.8$ Hz, H-3), 6.72 (1H, s, N-H), 7.15 (1H, s, ArH), 7.17 (1H, d, $J = 2.0$ Hz, ArH), 7.21 (2H, d, $J = 8.4$ Hz, ArH), 7.26 (1H, d, $J = 2.0$ Hz, ArH), 7.35 (3H, d, $J = 8.8$ Hz, ArH), 7.43 (2H, d, $J = 8.4$ Hz, ArH), 7.50 (2H, d, $J = 8.8$ Hz, ArH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 43.6, 60.7, 66.9, 81.2, 115.7, 118.0, 120.1, 122.2, 123.5, 128.6, 129.6, 129.8, 135.3, 150.7, 155.9, 160.7, 163.1, 164.7, 165.2, 170.7; GC-MS m/z 878.34 [M^+]; Analysis calculated for $\text{C}_{43}\text{H}_{47}\text{ClN}_{12}\text{O}_7$: C, 58.73; H, 5.39; N, 19.11%. Found: C, 59.04; H, 5.01; N, 19.77%.

4.9.21. 3-(2,4-dichlorophenoxy)-1-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)amino)phenyl)-4-(4-((4,6-dimorpholino-1,3,5-triazin-2-yl)oxy)phenyl) azetidin-2-one (12c)

White solid; Mp. 285–287 °C; IR (KBr, cm^{-1}) 2854, 1751 (CO β -lactam), 1612, 1573, 1504; ^1H NMR (400 MHz, CDCl_3) δ 3.75–3.78 (32H, m, $16 \times \text{CH}_2$), 5.42 (1H, d, $J = 4.4$ Hz, H-4), 5.57 (1H, d, $J = 4.4$ Hz, H-3), 6.88 (1H, s, NH), 6.95 (1H, t, $J = 7.2$ Hz, ArH), 7.13 (2H, d, $J = 8.4$ Hz, ArH), 7.19 (2H, t, $J = 7.6$ Hz, ArH), 7.35 (2H, d, $J = 8.8$ Hz, ArH), 7.40 (2H, d, $J = 8.0$ Hz, ArH), 7.49 (2H, d, $J = 8.4$ Hz, ArH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 43.7, 61.7, 66.8, 81.4, 115.7, 117.9, 120.1, 121.9, 122.50, 122.3 128.6, 129.3, 131.6, 136.3, 152.8, 155.9, 157.0 162.5, 164.1, 165.1, 165.9, 170.8; GC-MS m/z 912.30 [M^+]; Analysis calculated for $\text{C}_{43}\text{H}_{46}\text{Cl}_2\text{N}_{12}\text{O}_7$: C, 56.52; H, 5.07; N, 18.39%. Found: C, 56.84; H, 5.41; N, 18.97%.

4.10. DPPH radical-scavenging activity assay

For evaluating radical-scavenging activity, the diphenylpicrylhydrazyl (DPPH) scavenging assay was performed as described previously.^{58–60} Briefly, a suitable dilution of test compound (0.05 ml) (dissolved in DMSO) was mixed with a solution of DPPH in methanol ($A_{517} = 1.0$; 2.95 ml) and the UV absorbance at 517 nm was measured for 5 min. Percent radical scavenging was calculated as $100 \times (A_{\text{start}} - A_{\text{end}})/A_{\text{start}}$, where A_{start} is the absorbance before addition of test compound and A_{end} is the absorbance value after 5 min of reaction time.

4.11. Antiproliferative activity assay

The antiproliferative activity assay was carried out as described by Rowan et al.⁶⁸ with slight modifications. The HepG2 and SW 1116 cell lines were used in which the cell monolayers were seeded at 5×10^4 cells per well (in RPMI 1640 medium plus 10% fetal calf serum) in a 96-well plate. Various concentration of tested compounds (5, 10, 50, 100 and 200 μM) were prepared and inoculated into the cell monolayer and repeated in triplicate for each compound. Monolayers containing the tested compounds were incubated overnight at 37 °C in a 5% CO_2 atmosphere. After overnight incubation, the suspension from each well was discarded and 25 μl of fresh complete medium (RPMI+10% fetal calf serum) containing 0.004 g/ml of MTT reagent (Sigma, Ronkonkoma, NY, USA) was added. Samples were incubated for 3 h at 37 °C in 5% CO_2 and the formazan product was solubilized by the addition of 100 ml of DMSO. Optical densities of the suspensions were measured at 540 nm using an ELISA reader (Biotek, Power Wave, Winooski, VT, USA). The absorbance (optical density) was measured and percentage inhibition or percentage cell death was calculated. Doxorubicin was used as a reference positive control compound. The results of the MTT assay are expressed as mean IC_{50} values.

4.12. UV-Visible spectroscopy

Calf thymus DNA (CT-DNA) and all reagents and materials were purchased from Sigma-Aldrich. Experiments were done in 50 mM Tris-HCl buffer containing 100 mM NaCl, pH 7.4. CT-DNA stock solution was prepared by dissolving the solid DNA into doubly-distilled water overnight, and stored at 4 °C in the dark for no more than one week. The concentration of CT-DNA was determined by absorption spectrometry, using its known extinction coefficient at 260 nm ($6600 \text{ M}^{-1} \text{ cm}^{-1}$)⁷⁰. Solutions of CT-DNA in Tris-HCl buffer gave a ratio of UV absorbance at 260 and 280 nm (A_{260}/A_{280}) of 1.8–1.9, showing that DNA is sufficiently protein-free.⁵⁵ The stock solutions of compound **7b** (68 μM), **8g**, **8l**, and **8f** (40 μM) were prepared by dissolving their powder into suitable amounts of DMSO solutions. All spectroscopic measurements were done at 25 °C and 2 min after addition of the test compounds.

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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